**Cell Culture Medium**

**10% FBS supplemented DMEM**

**Description**

This is the cell culture medium

**Ingredients**

Add the following solutions to 100ml DMEM

- warm up and add 100% FBS 11 ml (10%) - from freezer
- penecilin/streptomyosin 1.1 ml (1%) – from freezer
- sodium pyruvate 1.1 ml (1%) – from fridge
- L-glutamine 1.1 ml (1%) – from freezer.

**10% FBS without penecilin/streptomyosin**

**Description**

This is the cell culture medium for transfection. It is the same as the cell culture medium, but without penecilin/streptomyosin.

**Ingredients**

Add the following solutions to 100 ml DMEM

- warm up and add 100% FBS 11ml (10%)
- sodium pyruvate 1.1ml (1%)
- L-glutamine 1.1ml (1%)

**0.5% FBS supplemented DMEM**

**Description**

This is the cell culture medium for starving cells

**Ingredients**

Add the following solutions to 100ml DMEM

- warm up and add 100% FBS 0.5 ml (0.5%)
penicillin/streptomycin 1.0 ml (1%)
sodium pyruvate 1.0 ml (1%)
L-glutamine 1.0 ml (1%)

0.5% FBS without penicillin/streptomycin

Description
This is the cell culture medium for starving cells, when the cell condition is not very good.

Ingredients
Add the following solutions to 100 ml DMEM

- warm up and add 100% FBS 0.5 ml (0.5%)
- sodium pyruvate 1.0ml (1%)
- L-glutamine 1.0ml (1%)

CO₂ independent medium with 0.5% FBS

Description
Change cell cultures to this medium right before imaging under microscope

Ingredients

- 49-50 ml CO₂ independent medium
- 1 ml L-glutamine (200 mM)
- 250 ul 100% FBS (0.5%)

0.5X trypsin in PBS (40ml)

- warm up and add 2 ml 10X trypsin-EDTA (in freezer) to 38 ml PBS

2% Gelatin (200 ml)

- get 200 ml MQ H₂O, weigh 4 g Gelatin (cell culture grade) and add to H₂O, gently shake bottle, put on a tray containing some water, loose the cap of the bottle, autoclave with liquid cycle (30min). After take out, be sure to shake the bottle until gelatin desolve uniformly in liquid, keep in refrigerator for later use.